

### OPERATION MODE

### SERVICE MODE

## 01 Use a priming plate for start up and shutdown priming

Prepare 500mL per day each of:  
1) 70% ethanol + 1% Tween 20  
then

2) DI with 1% Tween 20



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## 02 Switch to a calibration plate (change monthly)

Visually inspect the droplet after dispensing 80  $\mu$ L and again after three cycles to ensure even volumes

### SERVICE MODE



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Dispense 80 $\mu$ L inspect, then run three cycles and inspect again

### OPERATION MODE

- 10  $\mu$ L/s flow rate
- 80  $\mu$ L initial volume
- 3 cycles

## 03 Prime with assay washing buffer

### SERVICE MODE



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1. Select the Change Buffer button
2. Select the desired buffer channel to prime
3. Add the priming plate to the HT2000 tray and follow the onscreen instructions to initiate the buffer change and a prime

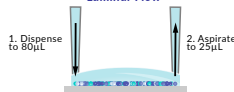
### OPERATION MODE

### SERVICE MODE

## 04 Running an assay

One wash cycle consumes 5.5mL buffer

Laminar Flow



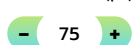
7 cycles = complete buffer exchange

Flow Rate ( $\mu$ L/s)



- 5 Maximize cell retention
- 10 Typical operation
- 20 Extreme wash (not recommended)

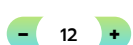
Initial Volume ( $\mu$ L)



Maximize cell settling: 25 - 75  $\mu$ L (Vortex-able at 25 - 50  $\mu$ L).

Wells can accommodate:  $\leq$ 3M cells  $\leq$ 75  $\mu$ L (140  $\mu$ L with Adapter)

Wash Number



- 7 Buffer exchange
- 12 Moderate wash
- 25 Extensive wash

\* If BX10 model is installed, ensure that the buffer configuration (Primary or Primary + Secondary) is correct before starting an assay

### OPERATION MODE

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## 05 When finished for the day

Use a priming plate for Shutdown Prime and Nozzle Prime



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- 1) Shutdown Prime
  - a) DI with 1% Tween 20 then
  - b) 70% ethanol + 1% Tween 20 then



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- 2) Nozzle Prime with 70% ethanol + 1% Tween 20
 

\*Note: the plate will remain half-full of 70% ethanol + 1% Tween 20 once Nozzle Prime finishes
- 3) Proceed with BX Maintenance
- 4) Power off

## 06 BX Maintenance

- 1) Connect the single side of the 1-to-4 splitter to DI + Tween 20 reservoir bottle
- 2) Connect the lines for EtOH, DI, WB-A and WB-B to the other end of the 1-to-4 splitter
- 3) Run BX Maintenance and select "Primary"
- 4) Repeat connected to the EtOH reservoir
- 5) Repeat disconnected to the reservoir to run air



\* If BX10 model is installed and lines for WB-C, WB-D, WB-E, or WB-F were used, please connect those to the splitter and repeat. If secondary channels were used, make sure to run BX Maintenance on "Secondary"

Is your SOP wash at 4°C

- Incubate on ice
- Keep buffers on ice during wash
- Keep instrument in the cold room

Ask your FAS about directly acquiring your samples from the Laminar Wash plate with your flow cytometer

For general settling times per cell count and volume, please refer to the New User Training Guide or contact your FAS



### Questions? Need Support?

Fill out our tech support form (serial number needed) or contact your FAS

